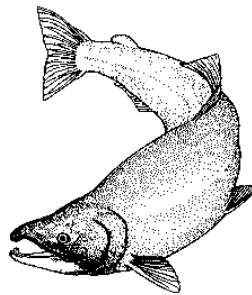


DRAFT

HATCHERY AND GENETIC MANAGEMENT PLAN
HGMP

Coho Salmon

Little White Salmon/Willard NFH Complex



U.S. Fish & Wildlife Service
Columbia River Fisheries Program Office
Little White Salmon/Willard National Fish Hatchery Complex

December 14, 1999

HATCHERY AND GENETIC MANAGEMENT PLAN

U.S. Fish & Wildlife Service

The purpose of this hatchery and genetic management plan (HGMP) template is to provide a single source of hatchery information for comprehensive planning by federal, state, and tribal managers, and for permitting needs under the Endangered Species Act (ESA).

Section 1. General Program Description

1.1) Name of Program: Early Run Coho Program -
Little White Salmon/Willard NFH Complex

1.2) Population (or stock) and species: Lower Columbia River Early Run Coho Salmon
(*Oncorhynchus kisutch*)

1.3) Responsible organization and individual:

Name(and title): Lee Hillwig (Fish and Wildlife Administrator)

Organization: U.S. Fish and Wildlife Service (Service)

Address: 911 N.E. 11th Avenue, Portland, Oregon 97232

Telephone: (503) 872) 2766

Fax: (503) 231-2062

Email: lee_hillwig@fws.gov

Other organizations involved, and extent of involvement in the program:

- National Marine Fisheries Service (NMFS) - funding agency via Mitchell Act.
- Nez Perce Tribe receives production for tribal restoration program.
- Yakama Indian Nation receives production for tribal restoration program.
- U.S. v Oregon parties - co-managers of fisheries.

1.4) Location(s) of hatchery and associated facilities:

Little White Salmon NFH is located on the Little White Salmon River at river kilometer 2, approximately 19 kilometers east of Stevenson, Washington. The hatchery is situated just above Drano Lake, a water body where the Little White Salmon River joins the Columbia River at river kilometer 261. Site elevation is about 27 meters above sea level. Willard NFH is located on the Little White Salmon River approximately 6.5 kilometers upstream from the Little White Salmon NFH. These two hatcheries are operated as the Little White Salmon/Willard NFH Complex (Complex) .

1.5) Type of program: Mitigation

1.6) Purpose (Goal) of program:

Little White Salmon River Program:

Little White Salmon NFH was originally constructed in 1898 and was remodeled and expanded in 1958. Willard NFH was authorized by the Mitchell Act in 1946 and constructed in 1952. The Complex currently operates as part of the Columbia River Fisheries Development Program under U.S. v Oregon and is funded through the Mitchell Act—a program to provide for the conservation of Columbia River fishery resources. The purpose is to successfully rear and release 1,000,000 locally adapted yearling coho salmon smolts for release on-station to help mitigate (production for fisheries) for fish losses in the Columbia River Basin caused by mainstem hydro-power project construction and other development, which will contribute to tribal commercial, ceremonial, and subsistence fisheries and non-tribal commercial and sport fisheries, while providing adequate escapement for hatchery production. Hatchery operations strive to meet mitigation requirements of the Mitchell Act and the Columbia River Fish Management Plan goals (U.S. v Oregon). The Columbia River Fish Management Plan is currently under renegotiation, however, current production goals are generally consistent with the production goals in the expired plan.

Clearwater Program:

The purpose is to rear coho salmon to be transferred to the Nez Perce Tribe and released into natural habitat of Lapwai and Potlatch Creeks to establish a naturally spawning, self-sustaining population to help restore and recover an extirpated stock of fish. As an interim goal, a total of 500,000 coho salmon are reared at the Little White Salmon/Willard National Fish Hatchery Complex using Mitchell Act funds. These fish receive coded-wire tags and passive integrated transmitter tags to help evaluate the success of this restoration and recovery effort. As specified in the Nez Perce Tribal Coho Master Plan, coho originating from the Complex are transported and released into natural habitat of Lapwai and Potlatch Creeks, two tributaries of the Clearwater River, Idaho, located on the Nez Perce Indian Reservation. Fish returning to these creeks will be allowed to spawn naturally to help recover a self-sustaining population of fish. The program began with the 1995 brood year, released in 1997. The interim goal is to develop a locally adapted brood stock from the Clearwater River system releases that would be used for rearing at the Complex for the Nez Perce restoration program. The Clearwater program is not evaluated in this HGMP. The Nez Perce Tribe is responsible for the transfer of the fish to the Clearwater River system and for evaluating the success of this program. It should be covered under a separate HGMP for the BPA funded Nez Perce tribal program.

Yakima Program:

The purpose is to rear and mark coho salmon for eventual release into natural habitat of the Yakima River to help restore this species to historic levels and evaluate the success of this restoration effort. A total of 1 million coho salmon are reared at the Little White Salmon/Willard National Fish Hatchery Complex using Mitchell Act funds. Fish are transferred to the Yakama Indian Nation (YIN). The fish are acclimated and released into the Yakima and Naches River Basins, located on the Yakama Indian Reservation, as part of a tribal restoration and research effort. These fish are 100% marked with coded wire tags using BPA funds. Unique tag codes

DRAFT

are used to evaluate the success of this restoration effort at three tribal acclimation sites located in the Yakima Basin. The first releases will be made into these three areas from the Little White Salmon/Willard National Fish Hatchery Complex during February and March 2000 to initiate the development of locally adapted, naturally spawning populations of fish. The interim goal is to develop a locally adapted brood stock from the Yakima River system that would be used for rearing at the Complex for the Yakama tribal program. The Yakima program is not evaluated in this HGMP. It should be covered under a separate HGMP for the BPA funded Yakama tribal program.

1.7) Specific performance objective(s) of program:

The following objectives are adapted from IHOT (1995).

- Objective 1: Hatchery Production
Produce 1 million coho smolts for on-station release.
Produce 1.5 million coho pre-smolts for transfer.

- Objective 2: Minimize interactions with other fish populations through proper rearing and release strategies.

- Objective 3: Maintain stock integrity and genetic diversity of each unique stock through proper management of genetic resources.

- Objective 4: Maximize survival at all life stages using disease control and disease prevention techniques. Prevent introduction, spread or amplification of fish pathogens.

- Objective 5: Conduct environmental monitoring to ensure that hatchery operations comply with water quality standards and to assist in managing fish health.

- Objective 6: Communicate effectively with other salmon producers and managers in the Columbia River Basin.

1.8) List of Performance Indicators designated by "benefits" and "risks":

Information is not required at this time and may be provided at a later date, per guidance by NMFS on October 5, 1999.

1.9) Expected size of program:

The following is a program summary adapted from IHOT (1996).

<u>Measures</u>	<u>Hatchery Goal</u>	<u>5-Year Average</u>	<u>Range</u>
Adult Capture ¹	2,500	2,614	502 - 8,435
Fish Releases ^{1, 2, 3}	1 Million	1.6M	0.7M - 2.4M

DRAFT

Egg Transfers ¹	0	0	
Fish Transfers ^{1, 4, 5}	1.5 Million	1.8M	0.7M - 2.7M
Adults Passed Upstream ¹	0	0	0
Percent Survival, Juvenile to Adult ^{3, 6}	0.2%	0.13%	0.10% - 0.24%
Smolt Size at Release (fish/lb) ¹	15	16.7	15.0 - 20.3

- ¹ Five year average and range from calendar years 1995-1999
- ² Through 1998 the on-station release goal was 2 million smolts
- ³ On-station releases only
- ⁴ Clearwater Program began in 1997
- ⁵ Yakima program begins in 2000
- ⁶ From completed brood years 1990-1994

1.10) Date program started or is expected to start:

The program began in 1960.

1.11) Expected duration of program:

Ongoing program.

1.12) Watersheds targeted by program:

Little White Salmon River Program:

The Little White Salmon River below Little White Salmon NFH (i.e. Drano Lake) is the target watershed. Willard NFH, the release point for the coho salmon reared at the Complex, is located at river kilometer 9 on the Little White Salmon River, entering the Columbia River at river kilometer 261. This position is approximately 45° 45' 45" North Latitude and 121° 37' 30" West Longitude (pers. comm. Steve Vigg, NMFS).

1.13) Future program direction:

The future direction of this program may change as regional decision makers address salmon and steelhead restoration needs. As changes occur in hydro, habitat and harvest, and as hatchery reform is implemented, adaptive management strategies may include redirection of this program.

As such changes occur, or where new information becomes available that may potentially effect listed salmon and steelhead species, the Service will reinitiate consultation by supplementing this HGMP.

Section 2. Relationship of Program to Other Management Objectives

2.1) List all existing cooperative agreements, memoranda of understanding, memoranda of agreement, or other management plans or court orders under which program operates. Indicate whether this HGMP is consistent with these plans and commitments, and explain any discrepancies.

The coho salmon program is consistent with:

- NMFS 1999 Biological Opinion on Artificial Propagation in the Columbia River Basin
- U.S. v Oregon Columbia River Fish Management Plan (currently under re-negotiation)
- 1999 Management Agreement for Upper Columbia River Fall Chinook, Steelhead and Coho (under U.S. v Oregon)
- Mitchell Act
- Nez Perce Tribal Coho Master Plan
- Mid-Columbia Tribal Coho Plan (under development)
- IHOT Policies and Procedures for Columbia Basin Anadromous Salmonid Hatcheries

This HGMP is consistent with these plans and commitments.

2.2) Status of natural populations in target area.

The backwater from Bonneville Dam covers all of the area that was originally suitable for salmon spawning in the Little White Salmon River (Bryant 1949, WDFW 1990). See Section 5.2.3 below.

2.2.1) Geographic and temporal spawning distribution.

2.2.2) Annual spawning abundance for as many years as available.

2.2.3) Progeny-to-parent ratios, survival data by life-stage, or other measures of productivity for as many brood years as available.

2.2.4) Annual proportions of hatchery and natural fish on natural spawning grounds for as many years as possible.

2.2.5) Status of natural population relative to critical and viable population thresholds.

2.3) Relationship to harvest objectives.

There is no natural spawning population of coho in the Little White Salmon River. The hatchery barrier dam, as well as an impassable falls just upstream, precludes access of anadromous species into the upper basin and there is virtually no natural spawning area for coho salmon below the hatchery in the river (see 5.2.3). Therefore all returning fish above brood stock needs are harvestable. All coho released on-station are mass marked (adipose clipped) for selective harvest

purposes. Complex coho contribute to ocean commercial and sport fisheries; in-river mainstem sport, commercial, and tribal fisheries; and terminal area sport and tribal fisheries. West coast ocean fisheries are managed to achieve Oregon coastal natural coho stock limitations with an exploitation rate less than 15 percent. The Salmon Technical Team projected an 8.7 percent exploitation rate for the Oregon coastal natural coho stock in 1999 fisheries (STT 1999). The State of Oregon recently listed Clackamas wild coho as an endangered species and instituted a harvest management cap of a 15 percent harvest rate, including ocean impacts. These weak stock management restrictions directed at other coho stocks along with jeopardy standard restrictions for Snake River wild fall chinook and wild Group B steelhead effectively keep coho fishery impacts at low levels relative to very high harvest rates in past fisheries. Therefore, production releases of Complex coho are not expected to add adverse effects to listed species or other stocks of concern from a harvest management perspective beyond those currently allowable under non-jeopardy biological opinions for harvest. The 1999 fall season harvest biological opinion determined that fisheries managed to stay within the Snake River wild fall chinook and wild Group B steelhead jeopardy standards would not jeopardize any of the other listed species (NMFS 1999a).

Detailed survival and contribution information for those brood years coded-wire tagged from 1981 through 1994 (last completely returned brood year) is in Appendix A1-A9 (from Pastor 1999). Percent survival during this period ranged from 0.072 percent (BY 1991) to 1.670 percent (BY 1988), and averaged 0.412 percent.

2.4) Relationship to habitat protection and recovery strategies.

The backwater from Bonneville Dam covers all of the area that was originally suitable for salmon spawning in the Little White Salmon River (Bryant 1949, WDFW 1990). See section 5.2.3 below. Habitat restoration for anadromous salmonids outside of the Little White Salmon River basin will be a long term effort. The Mitchell Act mitigation program is in place precisely because habitat was lost to natural spawning populations of coho salmon. If mitigation goals for lost and degraded habitat are to be achieved, continued hatchery production will be required.

2.5) Ecological interactions.

Salmonid and non-salmonid fishes or other species that could:

1) negatively impact program;

A variety of freshwater and marine predators such as northern pikeminnows, Caspian terns, and pinnipeds, can significantly reduce overall survival rates of program fish. Predation by northern pikeminnow poses a high risk of significant negative impacts on the productivity of hatchery chinook (SWIG 1984). Based on PIT tags recovered at a large Caspian tern nesting colony on Rice Island, a dredge material disposal island in the Columbia river estuary, 6-25 million of the estimated 100 million out-migrating juvenile salmonids reaching the estuary were consumed by the terns in 1997 (Roby, et al. 1997). The Fish Passage Center (Berggren 1999) estimates, from about 57,000 PIT tag recoveries from Rice Island, that through 1991, about 0.2% of all PIT tagged fish released into the

DRAFT

Columbia River showed up on Rice Island. That percentage had increased by a factor of ten by the 1997 and 1998 juvenile salmonid out-migrations, with hatchery and wild steelhead having been the most effected by the increased predation. A NMFS Working Group (NMFS 1997) determined that California sea lion and Pacific harbor seal populations in the three west coast states have risen by 5-7% annually since the mid-1970s. Their predation on salmonids may now constitute an additional factor on salmonid population declines and can effect recovery of depressed populations in some situations. See the ecological interactions discussion below.

2) be negatively impacted by program;

Co-occurring natural salmon and steelhead populations in local tributary areas and the Columbia River mainstem corridor areas could be negatively impacted by program fish. Of primary concern are the ESA listed endangered and threatened salmonids: Snake River fall-run chinook salmon ESU (threatened); Snake River spring/summer-run chinook salmon ESU (threatened); Lower Columbia River chinook salmon ESU (threatened); Upper Willamette River chinook salmon ESU (threatened); Upper Columbia River spring-run chinook salmon ESU (endangered); Columbia River chum salmon ESU (threatened); Snake River sockeye salmon ESU (endangered); Upper Columbia River steelhead ESU (endangered); Snake River Basin steelhead ESU (threatened); Lower Columbia River steelhead ESU (threatened); Upper Willamette River steelhead ESU (threatened); Middle Columbia River steelhead ESU (threatened); and the Columbia River distinct population segment of bull trout (threatened). An additional concern is the Southwestern Washington/Columbia River coastal cutthroat trout ESU proposed for listing as threatened. See the ecological interactions discussion below.

3) positively impact program;

Returning chinook and other salmonid species that naturally spawn in the target stream and surrounding production areas may positively impact program fish. Decaying carcasses may contribute nutrients that increase productivity of the overall system.

4) be positively impacted by program;

A host of freshwater and marine species that depend on salmonids as a nutrient and food base may be positively impacted by program fish. The hatchery program may be filling an ecological niche in the freshwater and marine ecosystem. A large number of species are known to utilize juvenile and adult salmon as a nutrient and food base (Groot and Margolis 1991; and McNeil and Himsworth 1980). Pacific salmon carcasses are also important for nutrient input back to freshwater streams (Cederholm et al. 1999). Reductions and extinctions of wild populations of salmon could reduce overall ecosystem productivity. Because of this, hatchery production has the potential for playing an important role in population dynamics of predator-prey relationships and community ecology. The Service speculates that these relationships may be particularly important (as either ecological risks or benefits) in years of low productivity and shifting climactic cycles.

DRAFT

In addition, wild co-occurring salmonid populations might be benefitted as schools of hatchery fish migrate through an area. The migrating hatchery fish may overwhelm predator populations, providing a protective effect to the co-occurring wild populations. See the ecological interactions discussion below.

The 1999 Biological Assessment for the Operation of Hatcheries Funded by the National Marine Fisheries Service under the Columbia River Fisheries Development Program (NMFS 1999b) and the 1999 Biological Opinion on Artificial Propagation in the Columbia River Basin (NMFS 1999c) present a discussion of the potential effects of hatchery programs on listed salmon and steelhead populations. The reader is referred to the discussion in those documents.

Nine generalized types of effects that artificial propagation programs can have on listed salmon and steelhead populations were identified. These effects include: 1. Hatchery operation, 2. Brood stock collection, 3. Genetic introgression, 4. Hatchery production (density-dependent), 5. Disease, 6. Competition, 7. Predation, 8. Residualism, and 9. Migration corridor/ocean. Potential effects in these categories may apply to all hatchery programs to one degree or another depending on the particular program design.

A discussion of ecological interactions relative to the Complex's coho on-station release program follows:

1. Hatchery operation- The water source for the Willard NFH is withdrawal from the Little White Salmon River. An impassable falls immediately upstream from the Little White Salmon NFH site in the lower Little White Salmon River precludes anadromous fish passage into the upper basin. Water withdrawals for hatchery operation do not impact listed anadromous species because there is essentially no natural spawning or rearing habitat accessible to anadromous species in the basin. Hatchery effluents meet established NPDES release standards criteria and are diluted by the flow in Little White Salmon River, reducing potential negative impacts to natural stocks.

2. Brood stock collection- Returning early stock coho are collected for brood stock at the Little White Salmon NFH rack near the mouth of the Little White Salmon River. Stray hatchery coho from other locations or returns from natural production are not known to occur at Little White Salmon NFH. Any unmarked fish returning to the facility are likely from Willard stock that escaped marking during the juvenile fin-clipping operation. Columbia River coho are currently not listed but natural populations continue to be a candidate species.

3. Genetic introgression- Complex coho are not known to contribute to a significant straying problem outside of the local area. There is essentially very little, if any, productive spawning habitat below Little White Salmon NFH at the mouth of the Little White Salmon River (Drano Lake). Historical coho habitat was inundated by Bonneville Pool when Bonneville Dam was constructed in 1938 (Bryant 1949).

DRAFT

4. Hatchery production (density dependent effects)- Willard NFH coho releases from the facility are moderate in magnitude (typically about 1.0 to 2.0 million coho smolts) relative to other Columbia River coho production programs. This level of release is not expected to cause serious density dependent effects in the mainstem Columbia River. All coho released on-station are marked (adipose clipped) to promote selective harvest while providing protection for wild stocks.

5. Disease- Hatchery programs routinely treat fish in response to disease outbreaks that occur, in part, because large numbers of fish are maintained under crowded conditions. Most pathogens now enter hatcheries through returning adult fish, surface water supplies, and other mechanisms involving direct contact with naturally spawning fish. Crowding and stress decrease the physiological resistance of salmonid fishes to disease and increase the likelihood of infection (Salonius and Iwama 1993; Schreck et al. 1993). Consequently, concern exists that the release of hatchery fish may increase the risk of disease in naturally spawning populations.

Fish managers largely understand the kinds, abundance and virulence (epidemiology) of pathogens and parasites in hatchery fish. Recent studies suggest that the incidence of some pathogens in naturally spawning populations may be higher than in hatchery populations (Elliot and Pascho 1994). Indeed, the incidence of high ELISA titers for *Renibacterium salmoninarum*, the causative agent of Bacterial Kidney Disease (BKD), appears, in general, to be significantly more prevalent among wild smolts of spring/summer chinook salmon than hatchery smolts (Congleton et al. 1995; Elliot et al. 1997). For example, 95% versus 68% of wild and hatchery smolts, respectively, at Lower Granite Dam in 1995 had detectable levels of *R. salmoninarum* (Congleton et al. 1995). Although pathogens may cause significant post-release mortality among hatchery fish, there is little evidence that hatchery origin fish routinely infect naturally produced salmon and steelhead in the Pacific Northwest (Enhancement Planning Team 1986; Steward and Bjornn 1990). Many biologists believe disease-related losses often go undetected, and that the impact of disease on naturally spawning populations may be underestimated (Goede 1986; Steward and Bjornn 1990). Nevertheless, we are unaware of any studies or documentation in the scientific literature where hatchery fish have infected a naturally spawning population of salmon or steelhead in the Pacific Northwest (see also Campton 1995).

The Complex follows Integrated Hatchery Operations Team (IHOT 1995) and Pacific Northwest Fish Health Protection Committee protocols for disease sampling and treatment. The Lower Columbia River Fish Health Center is located nearby at Spring Creek NFH so fish health sampling, diagnosis, and treatment is readily available as fish health issues arise. See section 10.4.3 for fish health details. The fish health goal for Willard NFH coho is to release healthy fish that are physiologically ready to migrate. Willard coho are released directly into the Little White Salmon River and pass only one mainstem Columbia River dam (Bonneville Dam) en route to the ocean. Complex coho have a much reduced potential for transmission of disease to other populations relative to other upriver programs which are subjected to the high density impacts and stresses of collection for transport and/or diversion through multiple bypass systems.

Our general conclusion at this time is that the Complex, as are all federal hatcheries in the

Columbia River Basin, is currently taking extensive measures to control disease and the release of diseased fish. As a consequence, infection of natural fish by hatchery fish does not appear to be a problem. Based on the relative prevalence of BKD among hatchery and wild chinook salmon (Elliot et al. 1997; Congleton et al. 1995), the crowding and handling of fish at transportation dams at the time of barging or bypass may have a greater likelihood of increasing the incidence of disease among naturally produced fish than direct infection from hatchery fish.

6. Competition- The impacts from competition are assumed to be greatest in the spawning and nursery areas at points of highest density (release areas) and diminish as hatchery smolts disperse (USFWS 1994). Salmon and steelhead smolts actively feed during their downstream migration (Becker 1973; Muir and Emmelt 1988; Sager and Glova 1988). Competition in reservoirs could occur where food supplies are inadequate for migrating salmon and steelhead. However, the degree to which smolt performance and survival are affected by insufficient food supplies is unknown (Muir and Coley 1994). On the other hand, the available data are more consistent with the alternative hypothesis that hatchery-produced smolts are at a competitive disadvantage relative to naturally produced fish in tributaries and free-flowing mainstem sections (Steward and Bjornn 1990). Although limited information exists, available data reveal no significant relationship between level of crowding and condition of fish at mainstem dams. Consequently, survival of natural smolts during passage at mainstem dams does not appear to be affected directly by the number - or density - of hatchery smolts passing through the system at present population levels. While smolts may be delayed at mainstem dams, the general consensus is that smolts do not normally compete for space when swimming through the bypass facilities (Enhancement Planning Team 1986). The main factor causing mortality during bypass appears to be confinement and handling in the bypass facilities, not the number of fish being bypassed.

Juvenile salmon and steelhead, of both natural and hatchery origin, rear for varying lengths of time in the Columbia River estuary and pre-estuary before moving out to sea. The intensity and magnitude of competition in the area depends on location and duration of estuarine residence for the various species of fish. Research suggests, for some species, a negative correlation between size of fish and residence time in the estuary (Simenstad et al. 1982).

While competition may occur between natural and hatchery juvenile salmonids in - or immediately above - the Columbia River estuary, few studies have been conducted to evaluate the extent of this potential problem (Dawley et al. 1986). The general conclusion is that competition may occur between natural and hatchery salmonid juveniles in the Columbia River estuary, particularly in years when ocean productivity is low. Competition may affect survival and growth of juveniles and thus affect subsequent abundance of returning adults. However, these are postulated effects that have not been quantified or well documented.

The release of hatchery smolts that are physiologically ready to migrate is expected to minimize competitive interactions as they should quickly migrate from the release site. The Complex's coho are released into the Little White Salmon River at the Willard NFH site and it is assumed that they migrate quickly into the mainstem Columbia River migration corridor en route to the

DRAFT

ocean reducing the potential for competitive interactions with listed stocks. Snorkel surveys, juvenile out-migrant traps, and/or PIT tagging would provide valuable information on the timing of emigration and level of residualism, but would require additional funding. There have been no mortalities recorded during saltwater challenges conducted during the last three brood years at the Complex. Released fish have been fully smolted and begin their downstream migration immediately following release. Because Complex coho releases occur “low” in the Columbia Basin system relative to many other upriver programs, there is reduced opportunity for competitive interactions.

Other observations leading to conclusions regarding the behavior of released smolts included physiological and survival data collected during recent NATURES rearing studies conducted for coho salmon at Willard NFH. For several brood years, researchers from the (now) Biological Resources Division of the U.S. Geological Survey collected data to evaluate the use of cover (simulating natural riparian cover) during hatchery rearing to improve the post-release survival of hatchery-reared coho salmon and to alter their behavior to more closely match wild (naturally produced) fish. Preliminary physiological and survival data collected to date indicate that, although there were no differences detected among treatment groups when compared to control groups, the behavior of hatchery-produced fish from the Complex appears to be normal when compared to naturally produced fish.

The information in Table 1 is indicative of the strong homing of Complex fish. These data were extracted from the Columbia River Information System (CriS) and Pacific States Marine Fisheries Commission (PSMFC) databases (Pastor 1999). Data listed in the table above show that straying, as a measure of homing ability, virtually did not occur. Refer to Appendix A for more details.

Table 1. Coded-wire tag recoveries from brood years 1991-1994.

Species	Brood Year	Where Recovered	Expanded No.
<u>Coho</u>	1991	ODFW Columbia R. Gillnet	491
		ODFW Freshwater Sport	61
		FWS Hatchery	1,657
	1992	ODFW Columbia R. Gillnet	161
		FWS Hatchery	1,486
		ODFW Ocean Sport	120
		WDFW Sport (charter)	120
		WDFW Sport (private)	120
	1993	ODFW Columbia R. Sport	85
		FWS Hatchery	733
		ODFW Hatchery	14
		WDFW Sport (charter)	28

DRAFT

	1994	ODFW Estuary Sport	285
		FWS Hatchery	4,558
		WDFW Sport (charter)	47
		WDFW Sport (private)	712

There are no natural fish populations that spawn in the target area. Fish headed further up the Columbia River may dip into Drano Lake and hold in the favorable water conditions. Characteristic of steelhead, this species holds in Drano Lake during periods of low Columbia River flow and high water temperature, preferring the cooler Little White Salmon River water during the period of July through August. This period is sooner than the return migration of coho entering the adult holding ponds at the Complex. It is doubtful that there is any negative interaction between program fish and any natural fish.

7. Predation- The Complex's releases of coho occur in the upper Little White Salmon River at Willard NFH where other anadromous stocks do not have access. Predation effects would therefore be limited to the migration corridor where effects are likely to be reduced relative to spawning and nursery areas. Depending on species and population, hatchery smolts are often released at a size that is greater than their naturally-produced counterparts. In addition, for species that typically smolt at one year of age or older (e.g. steelhead, spring chinook salmon), hatchery-origin smolts may displace younger year classes of naturally-produced fish from their territorial feeding areas. Both factors could lead to predation by hatchery fish on naturally produced fish, but these effects have not been extensively documented, nor are the effects consistent (Steward and Bjornn 1990). The USFWS (1994) presented information that salmonid predators are generally thought to prey on fish approximately one-third or less their size.

The southwestern Washington/Columbia River coastal cutthroat trout ESU is proposed for listing as threatened under the ESA. While there is no population of coastal cutthroat trout in the Little White Salmon River, program fish from the Complex could potentially encounter out-migrants of the sea-run form of the cutthroat in the mainstem or estuary of the Columbia River. Trotter (1997) states that the time of seaward movement of sea-run cutthroat in Oregon and Washington streams begins as early as March and peaks in mid-May. In some lower Columbia River tributaries in Washington, the USFWS found a similar run timing as presented in Trotter (1997) and also found that the size of the sea-run cutthroat trout smolts ranged from 100mm-260mm (S. Barndt, USFWS, pers. comm.). Sea-run cutthroat out-migrants are very similar in timing and size to the yearling hatchery smolts released from the Complex. Instances of predation by hatchery smolts are thought to be low.

In general, the extent to which salmon and steelhead smolts of hatchery origin prey on fry from naturally reproducing populations is not known, particularly in the Columbia River basin. The available information - while limited - is consistent with the hypothesis that predation by hatchery-origin fish is, most likely, not a major source of mortality to naturally reproducing

DRAFT

populations, at least in freshwater environments of the Columbia River basin (Enhancement Planning Team 1986). For example, peak emergence of listed chum salmon at Ives Island, a natural production area below Bonneville Dam, was estimated to occur during the latter half of March in 1999 (2/19/99 fax to Donna Allard from Wayne Vander Naald, ODFW). Out-migrant sampling conducted by the USFWS in 1998 and 1999 in Hardy Creek, which is adjacent to the mainstem Pierce/Ives Island natural production area, indicated that peak emigration of chum fry from this tributary occurred during the first two weeks of March (unpublished data). Based on life history traits, it is expected that most of the chum fry would have emigrated from the natural production area before the mid-April release of larger hatchery coho occurs at the Complex. The potential for the Complex smolts to prey on emerging chum fry would not be significant. However, virtually no information exists regarding the potential for such interactions in the marine environment.

The presence of large numbers of hatchery fish may also alter the listed species behavioral patterns, which may influence vulnerability and prey susceptibility (USFWS 1994). Releasing large numbers of hatchery fish may also lead to a shift in the density or behavior of non-salmonid predators, thus increasing predation on naturally reproducing populations. Conversely, large numbers of hatchery fish may mask or buffer the presence of naturally produced fish, thus providing sufficient distraction to allow natural juveniles to escape (Park 1993). Prey densities at which consumption rates are highest, such as northern pikeminnow in the tailraces of mainstem dams (Beamesderfer et al. 1996; Isaak and Bjornn 1996), have the greatest potential for adversely affecting the viability of naturally reproducing populations, similar to the effects of mixed fisheries on hatchery and wild fish. However, hatchery fish may be substantially more susceptible to predation than naturally produced fish, particularly at the juvenile and smolt stages (Piggins and Mills 1985; Olla et al. 1993).

Predation by birds and marine mammals (e.g. seals and sea lions) may also be significant source of mortality to juvenile salmonid fishes, but functional relationships between the abundance of smolts and rates of predation have not been demonstrated. Nevertheless, shorebirds, marine fish, and marine mammals can be significant predators of hatchery fish immediately below dams and in estuaries (Bayer 1986; Ruggerone 1986; Beamish et al. 1992; Park 1993). Unfortunately, the degree to which adding large numbers of hatchery smolts affects predation on naturally produced fish in the Columbia River estuary and marine environments is unknown, although many of the caveats associated with predation by northern pikeminnow in freshwater are true also for marine predators in saltwater.

8. Residualism- Complex coho releases are not known to residualize in the Little White Salmon River. Even if Willard coho do residualize there would be no effect on listed anadromous species because there is no access to anadromous species in this area. Snorkel surveys, out-migrant traps, and/or PIT tagging would help to provide a definitive answer to hatchery out-migration questions. This would require additional funding.

9. Migration corridor/ocean- The hatchery production ceiling called for in the Proposed Recovery

Plan for Snake River Salmon of approximately 197.4 million fish (1994 release levels) has been incorporated by NMFS into their recent hatchery biological opinions to address potential mainstem corridor and ocean effects as well as other potential ecological effects from hatchery fish. Although hatchery releases occur throughout the year, approximately 80 percent occur from April to June (NMFS 1999b) and Columbia River out-migration occurs primarily from April through August. The Complex's coho production is typically released in April, at the beginning of the general out-migration season for other hatchery and natural populations. The total number of hatchery fish released in the Columbia River basin has declined by about 26 percent since 1994 (NMFS 1999a) reducing potential ecological interactions throughout the basin.

Ocean rearing conditions are dynamic. Consequently, fish culture programs might cause density-dependent effects during years of low ocean productivity, especially in nearshore areas affected by upwelling (Chapman and Witty 1993). To date, research has not demonstrated that hatchery and naturally produced salmonids compete directly in the ocean, or that the survival and return rates of naturally produced and hatchery origin fish are inversely related to the number of hatchery origin smolts entering the ocean (Enhancement Planning Team 1986). If competition occurs, it most likely occurs in nearshore areas when (a) upwelling is suppressed due to warm ocean temperatures and/or (b) when the abundance or concentration of smolts entering the ocean is relatively high. However, we are only beginning to understand the food-chain effects of cyclic, warm ocean conditions in the eastern north Pacific Ocean and associated impacts on salmon survival and productivity (Beamish 1995; Mantua et al. 1997). Consequently, the potential for competition effects in the ocean cannot be discounted (Emlen et al. 1990).

Section 3. Water Source

Water rights total 33,868 gpm from the Little White Salmon River and springs. Water use for fish production ranges from 11,221 gpm to 28,232 gpm. The river supplies most of this water flow. The water intake structure was rebuilt in 1994. A water re-use system was constructed in 1967 for egg incubation. An independent hatchery audit (Montgomery Watson 1997) measuring hatchery operations against IHOT standards (IHOT 1995) reported a remedial action was needed to provide disease-free water for incubation and early rearing (4,700 gpm). The estimated cost was \$2.7 million. Such a system would also benefit the incubation and early rearing of upriver bright fall chinook and the incubation of coho.

The Complex's water intake structure was examined during the independent audit (Montgomery Watson 1997). The structure was in compliance when measured against NMFS's screening criteria for approach velocity and screen openings at that time. Subsequently, the screening criteria have been updated by NMFS and the structure has not been evaluated against the new criteria. If the structure is now out of compliance with the current screening criteria, there would be no impact on listed, or proposed to be listed, species. The intake structure is located well above the hatchery barrier dam. There are no reproducing populations of bull trout (*Salvelinus confluentus*) in the Little White Salmon River watershed (WDFW 1997).

Section 4. Facilities

Information is not required at this time and may be provided at a later date, per guidance by NMFS on October 5, 1999.

Section 5. Origin and Identity of Brood Stock

5.1) Source.

On-station releases into the Little White Salmon River:

- Adult coho salmon returning to the Little White Salmon River. Any “Early Run” coho is an acceptable sources of eggs or fish if shortfalls in adult returns occur.

Clearwater Program:

- Adult coho salmon returning to the Little White Salmon River. Any “Early Run” coho is an acceptable sources of eggs or fish if shortfalls in adult returns occur. The goal is to use locally adapted brood stock returning to the Clearwater River system for this program.

Yakima Program:

- Adult coho salmon returning to the Little White Salmon River. Any “Early Run” coho is an acceptable sources of eggs or fish if shortfalls in adult returns occur. The goal is to use locally adapted brood stock returning to the Yakima River system for this program.

5.2) Supporting Information.

5.2.1) History.

Initial attempts to rear coho salmon with the native, late running stock were made in 1919 and 1922 (Nelson and Bodle 1990). Attempts during the period 1930-1950 included the use of early run stock from the Quinault, Quilcene, Dungeness and Toutle Rivers. The Toutle River stock was considered responsible for establishing a successful run in 1956. By 1965, a dependable run of Toutle River coho had been established.

The following list contains facilities (brood source) that provided early-run coho eggs and/or fish for rearing at Willard NFH during the last 5 brood years:

- Lower Kalama Hatchery, WA
- Cascade Hatchery, OR
- Bonneville Hatchery, OR

- Speelyai Hatchery, WA
- Eagle Creek NFH, OR

5.2.2) Annual Size.

Adult coho salmon enter the hatchery holding ponds from late September through early November. Spawning occurs from mid-October to mid-November. A summary of total returns and numbers spawned from 1980 through 1999 is found in Appendix B. Adult returns ranged from 502 to 30,589, averaging 7,036, for this period. The annual escapement goal is 2,500 adults returning to the hatchery (see Section 1.9).

5.2.3) Past and proposed level of natural fish in brood stock.

As stated in Bryant (1949), the backwater from Bonneville Dam covers all of the area that was originally suitable for salmon spawning. In addition, a natural waterfall located about 0.8 kilometers above the hatchery barrier dam (built in 1974) had historically blocked access to spawning habitat located above the hatchery. Fluctuations in the level of the Bonneville Pool are seen immediately below the barrier dam. Historical literature reviews indicate that the only original native stock were the tule fall chinook and late-run coho (Nelson and Bodle 1990). Both are extinct from the watershed and there are no naturally spawning populations. There has been no past or proposed future level of natural fish used as brood stock for the coho salmon currently produced at the Little White Salmon/Willard NFH Complex.

5.2.4) Genetic or ecological differences.

As stated in section 5.2.3 above, there are no natural stocks in the Little White Salmon River.

5.2.5) Reason for choosing.

All stocks of coho were chosen due to their availability and the fact that they were a lower river early run variety of coho. All natural stocks of coho within the Little White Salmon River watershed had become extinct prior to 1938.

5.2.6) Unknowns.

Extinction of natural stocks and the current practice of managing brood stocks by large geographic regions (other than on a watershed basis, e.g. mid-Columbia Brights, early-run coho, Carson-stock spring chinook, etc.) has led to decisions effecting choice of brood stock. Although not endemic to the Little White Salmon River watershed, one would expect local adaptation of the existing hatchery stocks over time.

Section 6. Brood Stock Collection

Brood stock collection practices are consistent with the guidelines established by IHOT (1995).

6.1) Prioritized goals.

1. Collect an adequate number of adult fish at the Little White Salmon/Willard NFH Complex to achieve the following production goals:
 - 1,000,000 yearling coho salmon released on site.
 - 500,000 yearling coho salmon released off site in the Clearwater River, Idaho for the Nez Perce Tribe. This Mitchell Act-funded restoration effort has been implemented to restore an extinct stock of coho salmon to the Clearwater River Basin.
 - 1,000,000 coho salmon released off site on the Yakima Indian Reservation as part of a restoration effort to help restore this species to historic levels.

2. For all coho collected at the Little White Salmon/Willard NFH Complex, collect enough adult fish to assure a 1:1 spawning ratio.

3. For all coho collected at the Little White Salmon/Willard NFH Complex, operate the hatchery fish ladder to assure collection of fish for brood stock is representative of the entire spectrum of the run.

4. For all coho collected at the Little White Salmon/Willard NFH Complex, treat as needed with formalin to control fungus and maintain pre-spawning mortality below 2.5%.

6.2) Supporting information.

The following information is based on historical data and updated annually as new brood year information is collected.

6.2.1) Proposed number of each sex.

The annual escapement goal is 2,500 adults returning to the hatchery. The expected sex ratio and eggs per female are as follows:

% Male	% Female	Eggs/Female	Spawning Ratio	No. Males Needed	No. Females Needed
47.5	52.5	2,600	1:1	1,187	1,313

Not all females are spawned, and not all eggs taken result in released smolts. There is up to 5 percent adult pre-spawn mortality, up to 17 percent mortality from green egg to egg eye-up, up to 1.5 percent mortality from eye-up to ponding, and up to 8 percent mortality from ponding to smolt release.

6.2.2) Life-history stage to be collected (e.g. eggs, adults, etc.).

The coho production program is derived from the collection of adult fish returning to the

Little White Salmon/Willard NFH Complex.

6.2.3) Collection or sampling design.

All fish production for the Complex is initiated by adult collection at Little White Salmon NFH. An impassable natural waterfall, located approximately 0.8 kilometers upstream of the Little White Salmon facility prevents adult passage to Willard NFH.

Returning adult fish migrate through Drano Lake (backwater of the Bonneville Pool at the mouth of the Little White Salmon River) and up the Little White Salmon River, before entering the hatchery ladder. To facilitate and maximize adult collection, further migration is prevented by a concrete barrier dam. Constructed in 1974, the fish ladder and barrier dam were built in anticipation of new peaking levels at Bonneville Dam (USFWS 1987). River water is supplied to 2- 30' wide X 90' long X 6' deep adult holding ponds. Water exiting the ponds, in addition to a separate attraction water intake, supplies water to the fish ladder. Adult fish migrating up the ladder enter the ponds through a finger weir.

The collection of coho occurs concurrently with the collection of upriver bright fall chinook. Ladder operations begin during the third week of September. Historical records indicate that coho are the first fish collected and that an earlier ladder opening results in the collection of stray tule fall chinook from Spring Creek NFH. Upriver bright fall chinook begin their upstream migration in the Little White Salmon River later than coho, with the first fish collected near mid-October. Migration of both species occurs in extremely large numbers in some years, and it is not uncommon to collect more than 500 adult fish in a 24 hour period. The hatchery ladder is operated until maximum densities are achieved. If this occurs, the ladder is closed until excess fish are randomly removed from the ponds or spawning occurs. The ladder is then reopened to continue collecting adults from the full spectrum of the return run. Generally, the hatchery ladder is closed by mid-November at which time very few fish remain below the hatchery barrier dam.

6.2.4) Identity.

There are no naturally spawning populations of coho salmon within the Little White Salmon River watershed. As a result, differential marking is not required. However, all coho salmon juveniles released into the Little White Salmon River are marked with an adipose clip for the purpose of selective fisheries management. This mark is used to distinguish hatchery fish in a mixed stock fishery. The collection of an un-marked (non-adipose clipped) coho salmon does not equate to the collection of a “wild” fish. The recovery of an un-marked coho is most likely a member of the double index mark (CWT but not adipose clipped, see section 9.7) fish that escaped the on-station mass marking. Over half of the Complex coho production program is left unmarked and designated for off-site transfer to tribal acclimation and release areas. Strays from this group may contribute to unmarked fish returning to the Complex. All adult fish that receive coded wire tags for stock assessment are sampled and the tags retrieved. Tag codes are determined and reported following the spawning season.

6.2.5) Holding.

The holding period for coho salmon is very short and uneventful. An aluminum bar-grader is installed between the two adult holding ponds to allow segregation by size. It is important to attempt to separate the large fall chinook from the smaller coho salmon. The common crowding of these fish normally results to damage in the smaller coho, being most evident by increases in broken eggs and bloody females. The Complex goal for all species is to achieve a 2.5% or less pre-spawning mortality rate during the holding period.

6.2.6) Disposition of carcasses.

Coho salmon are not chemically treated during spawning. Carbon dioxide is used to induce anesthesia. These fish are fit for human consumption. First priority for excess and spawned carcasses is provided to the Yakama Indian Nation ceremonial and subsistence program. All other excess carcasses are processed by contractors for the U.S. Department of Justice, Federal Prisons Program. Those fish treated with formalin are held for a minimum of 5 days prior to the excess process. Following any erythromycin treatment, all fish are either rendered using a commercial based rendering company or buried on-station.

6.3) Unknowns.

There are no data gaps that have lead to uncertainties about brood stock collection.

Section 7. Mating

7.1) Selection Method.

As mentioned in section 6.2.3, brood stock are collected that represent the entire spectrum of the run. Fish are sorted and ripe females spawned until 100% of the fish have been checked. Green females are passed back to the holding ponds with an adequate number of males to assure a 1:1 mating ratio. The eggs collected during a given sort are considered an egg “take”. Male spawners are randomly selected during the sort. Jack males are used in proportions representative of their return rate. In years of high jack returns, a larger proportion of jacks are used as spawners up to a five percent maximum. The number of jacks to be spawned on a given day is subjectively defined by hatchery staff and is subject to jack availability and ripeness. After all of the adult fish being held have been sorted once and ripe females spawned, a maximum one week period is allowed to pass before the fish are re-sorted and newly ripened females spawned. The objective is to achieve maximum fertilization by spawning fish soon after ovulation and yet avoid the needless handling of green females. The re-sorting process continues until all fish are spawned. Since there are no naturally spawning coho salmon in the watershed, differentiating spawners based on natural stock origin from within the watershed is not a criteria.

7.2) Males.

If the hatchery escapement goal is met, then a 1:1 spawning ratio will be achieved. Achieving this

spawning ratio is one of the highest brood stock program goals at the Complex. During low escapement years, males have been re-used on an as-needed basis to maximize the total number of females available to spawn. In low escapement years it is better to spawn the available females (and not lose that genetic material), than discard them. Under these conditions, reusing male fish does not compromise the genetic diversity of the hatchery stocks. It was determined that, in all instances, a minimum escapement need had been met to maintain genetic diversity, although some male fish had to be reused to achieve production goals.

7.3) Fertilization.

It is important to note that at no time in the recent past has the Complex pooled the eggs of females prior to fertilization. Again, as mentioned in section 7.2 above, an intense effort is made to achieve a 1:1 spawning ratio. The following is a detailed description of the spawning protocol.

Adults are crowded from holding ponds and anesthetized using carbon dioxide. Anesthetized adults are then sexed and checked for ripeness. Ripe adults are killed with a blow to the head. Tails of all females spawned are cut to allow bleeding for approximately 3-5 minutes. Eggs are then removed using a Wyoming knife and collected in iodophor-disinfected colanders to drain ovarian fluid. The eggs are then transferred to iodophor-disinfected stainless steel buckets and sperm is added directly to the eggs. A 1:1 random spawning ratio is maintained and male jacks are used proportionally to their percentage of the run. The buckets containing eggs and sperm of individual (paired) fish are then transferred to the Little White Salmon hatchery nursery building (0.3 kilometers away) where water is added to activate the sperm. This process takes from 5-10 minutes. The fertilized eggs are stirred and allowed to rest for a minimum of thirty seconds, then washed and water hardened for one half hour in a 75 mg/L iodophor solution in individual Heath incubator trays. The eggs are incubated using single pass spring or well water.

Aseptic procedures are followed to assure the disinfection of equipment throughout the egg handling process. Samples are collected by fish health specialists to determine the incidence of infectious hematopoietic necrosis (IHN), erythrocytic inclusion body syndrome (EIBS), *Ceratomyxa shasta*, and screening for other pathogenic bacteria. Refer to section 10.4.3 for more fish health details.

Coho salmon are not sampled for *Renibacterium salmoninarum*. Segregation and culling by titre group is not performed since bacterial kidney disease is not a chronic problem with this species.

7.4) Cryopreserved gametes.

Cryopreservation of gametes is not performed at the Little White Salmon/Willard NFH Complex.

Section 8. Rearing and Incubation

Information is not required at this time and may be provided at a later date, per guidance by NMFS on October 5, 1999.

Section 9. Release

9.1) Life history stage, size, and age at release.

Size at release information is provided for coho released from the Complex directly into the Little White Salmon River from 1990 through 1999 in Appendix C. The last fingerling (less than one year old) release was in 1992. Except where noted as fingerling releases, the yearling (18 months old) releases have been consistent in size at release for nine of the last 10 generations, at about 16 fish per pound.

9.2) Life history stage, size and age of natural fish of same species in release area at time of release.

As reported in sections 2.2 and 5.2.5, there are no naturally produced anadromous salmonid fish species in the Little White Salmon River watershed.

9.3) Dates of release and release protocols.

For the last three generations, yearling coho salmon have been released during the third week of April. The exact day of the week has varied in an attempt to maximize survival by planning releases on known spill days at Bonneville Dam. All releases are forced.

9.4) Location(s) of release.

Coho yearlings are released at the Willard facility, approximately 7 kilometers above the Little White Salmon barrier dam.

9.5) Acclimation procedures.

Since coho yearlings destined for on-station release are reared using pass through water originating from the Little White Salmon River, acclimation procedures are not required.

9.6) Number of fish released.

Release information is provided for fish released from the Complex directly into the Little White Salmon River from 1990 through 1999 in Appendix C. Yearling smolt releases during this period ranged from 706,032 (BY 1993) to 3,063,615 (BY 1991), and averaged 2,015,542 coho salmon. Refer also to section 1.9.

9.7) Marks used to identify hatchery adults.

All hatchery produced coho destined for on-station release receive an adipose clip (mass mark) to assist with selective fishery management strategies in mixed stock fisheries. A double index mark (using a combination of marked and unmarked fish with coded wire tags) is used to evaluate coho salmon survival. Coded-wire tagging is performed as part of the Complex's stock assessment marking program.

Section 10. Monitoring and Evaluation of Performance Indicators

10.1) Marking.

As stated in section 9.7, an adipose clip is used to designate the fish as originating from the hatchery. Coded-wire tagging is performed as part of the Complex’s stock assessment program. A double index mark (using a combination of marked and unmarked fish with coded wire tags) is used to evaluate coho salmon survival.

The following table summarizes the Complex’s stock assessment marking program to date:

<u>Species</u>	<u>Release No.</u>	<u>No. Marked</u>	<u>Mark</u>
Coho	1,900,000*	1,900,000*	Ad
	50,000	50,000	Ad-CWT
	50,000	50,000	CWT

* The Complex is currently reducing its on-station release program from 2 million to 1 million smolts to provide more smolts for tribal restoration programs. With a 1 million smolt on-station release, 900,000 smolts would be mass marked with an adipose clip.

10.2) Genetic Data.

Information is not required at this time and may be provided at a later date, per guidance by NMFS on October 5, 1999.

10.3) Survival and Fecundity.

Information is not required at this time and may be provided at a later date, per guidance by NMFS on October 5, 1999.

10.4) Monitoring of performance indicators in Section 1.8.

10.4.1) Proportions of hatchery spawners in natural populations in target area.

As reported in sections 2.2 and 5.2.5, there are no natural anadromous salmonid populations located in the Little White Salmon River watershed. Washington Department of Fish and Wildlife conducts spawning ground surveys in the local tributary systems (Wind River, Little White Salmon River, Big White Salmon River, Klickitat River). Recovered CWT data are reported annually.

10.4.2) Ecological interactions between program fish and natural fish (same and other species) in target area.

In-stream ecological interaction studies need to be developed and projects funded. These should compliment projects under section 10.4.6

10.4.3) Disease control in the hatchery, and potential effects on natural populations.

DRAFT

Aseptic procedures are followed to assure the disinfection of equipment throughout the egg handling process (see section 7.3). Fish health samples are collected to determine the incidence of infectious hematopoietic necrosis (IHN), erythrocytic inclusion body syndrome (EIBS), other reportable viruses, *Ceratomyxa shasta*, and pathogenic bacteria. Other contributions to improved fish health in the coho salmon at the Complex include: maintaining optimal rearing densities, two prophylactic antibiotic feedings to yearlings, routine monthly fish health examinations, and formalin treatments on an as-needed basis to control external parasites and fungal infections.

The following procedures are in place to monitor and maintain the health of the fish at the Complex:

General Fish Health Monitoring

- After fish are hatched, a 60 fish sample is examined for reportable viruses.
- On at least a monthly basis, both healthy and clinically diseased fish from each fish lot are given a health exam. The sample includes a minimum of 10 fish per lot.
- At spawning, a minimum of 150 ovarian fluids and 60 kidney/spleens are examined for viral pathogens from each species.
- Prior to transfer or release, fish are given a health exam. This exam may be in conjunction with the routine monthly visit. This sample consists of a minimum of 60 fish per lot.
- Whenever abnormal behavior or mortality is observed, the fish health specialist will examine the affected fish, make a diagnosis and recommend the appropriate remedial or preventative measures.
- Reporting and control of specific fish pathogens are conducted in accordance with the Co-Managers Salmonid Disease Control Policy and the USFWS Fish Health Policy and Implementation Guidelines.

Fish and Egg Movements

- Movements of fish and eggs are conducted in accordance with the Co-Managers Salmonid Disease Control Policy and the USFWS Fish Health Policy and Implementation Guidelines.

Therapeutic and Prophylactic Treatments

- At spawning, eggs are water-hardened in iodophor as a disinfectant.
- Juvenile fish are administered antibiotics orally when needed for the control of

bacterial infections.

- Formalin (37% formaldehyde) is dispensed into water for the control of fungus on eggs and the control of parasites on juveniles and adult salmon. Treatment dosage and time of exposure varies with species, life-stage and condition being treated.
- Therapeutants approved by the U.S. Food and Drug Administration or those under Investigative New Animal Drug permits are used for treatments. Under special circumstances, extra-label usage of other animal drugs may be prescribed by a veterinarian to control resistant disease organisms.

Sanitation

- All eggs brought to the facility are surface-disinfected with iodophor as per the USFWS Fish Health Policy.
- All equipment (nets, tanks, rain gear) is disinfected with iodophor between different fish/egg lots.
- Different fish/egg lots are kept in separate ponds or incubation units. Water is not reused.
- Tank trucks or tagging trailers are disinfected when brought onto the station. Foot baths containing iodophor are strategically located on the hatchery grounds (i.e., entrance to hatchery building) to prevent spread of pathogens.

All of the above practices would minimize potential negative effects on natural populations of fish by lessening the chance for horizontally transmitted diseases when encountering Complex fish in the migration corridor or in the ocean.

10.4.4) Behavior (migration, spawning, etc.) of program fish.

- Time of migration and spawning of coho salmon at the Complex is tracked to determine if any noticeable shift in run and spawn timing is occurring
- Immediately prior to release, a representative sample of 100 fish are subjected to a 24 hour saltwater challenge at a salinity concentration of 30 parts per thousand (3‰). Observations regarding visibility of parr marks, coloration and overall survival are recorded and used to evaluate the degree of smoltification.

10.4.5) Homing and straying rates for program fish.

Coded-wire tag recovery data are used to document straying and homing rates of program fish.

- A minimum of one marked group of fish (CWT and adipose fin clipped) for each production group is released. Release information is reported to the Pacific States

- Marine Fisheries Commission (PSMFC) coast wide database.
- Heads from all marked returns to the Complex are recovered during spawning operations.
- CWT recovery data at the Complex is compiled and reported to the PSMFC coast wide database. CWT recovery data from various ocean and freshwater fisheries, stream spawning ground surveys, and other hatcheries are reported to the PSMFC coast wide database by the recovering agency.
- Estimates of survival, distribution, and contribution for Complex released fish are summarized in an Annual Stock Assessment report. Data from off site recoveries of Complex released fish, downloaded from the PSMFC coast wide database, are used in the analysis.

10.4.6) Gene flow from program fish into natural populations.

As mentioned in several previous sections, there are no natural anadromous salmonid populations located in the Little White Salmon River watershed. As a result, gene flow from program fish into natural populations within this watershed is not a concern.

A systematic program to annually monitor baseline genetic data of the fish produced at the Complex needs to be developed and funded. This genetic monitoring would include the use of DNA (e.g. microsatellite) markers and evaluation of life history characters (e.g., run timing, age, and size class distribution of adults). For example, the use of DNA markers could entail the sampling and analysis of approximately 50-75 adults each from the early, middle, and late spawn groups, at least initially. At a minimum cost of \$50 per fish, the overall cost of initializing such a genetic monitoring program for the hatchery spawners alone would be at least \$10,000 per stock. A genetic database for Complex production would provide needed information within the Complex to monitor the genetic traits and viability of the stock produced. Genetic profile comparisons between carcasses and naturally produced juveniles, with DNA markers, is highly desired. The information would be available to compare to natural stocks in local tributary systems to monitor any introgression or ecological interactions between program fish and natural fish (section 10.4.2).

Section 11. Research

Information is not required at this time and may be provided at a later date, per guidance by NMFS on October 5, 1999.

Section 12. Attachments and Citations

Literature Cited

Bayer, R.D. 1986. Seabirds near an Oregon estuarine salmon hatchery in 1982 and during the

DRAFT

- 1983 El Nino. Fish. Bull. 84: 279-286.
- Beamesderfer, R.C.P., D.L. Ward, and A.A. Nigro. 1996. Evaluation of the biological basis for a predator control program on northern squawfish (*Ptychocheilus oregonensis*) in the Columbia and Snake rivers. Canadian Journal of Fisheries and Aquatic Sciences 53: 2898-2908.
- Beamish, R.J. (ed.). 1995. Climate Change and Northern Fish Populations. National Research Council of Canada. Ottawa, Canada.
- Beamish, R.J., B.L. Thomson, and G.A. Mcfarlane. 1992. Spiny Dogfish Predation on Chinook and Coho Salmon and the Potential Effects on Hatchery-Produced Salmon. Transactions of the American Fisheries Society 121: 444-455.
- Becker, C.D. 1973. Food and growth parameters of juvenile chinook salmon, *Oncorhynchus tshawytscha*, in central Columbia River. Fish. Bull. 71: 387-400.
- Berggren, T. 1999. Updated analysis of PIT tags detected in bird guano on Rice Island. Memo to U.S. Fish and Wildlife Service. Fish Passage Center, Portland, Oregon.
- Bryant, F.G. 1949. A survey of the Columbia River and its tributaries with special reference to its fishery resources; Washington streams from the mouth of the Columbia River to and including the Klickitat River (Area I). U.S. Fish & Wildlife Service, Washington, D.C.
- Campton, D.E. 1995. Genetic effects of hatchery fish on wild populations of Pacific salmon and steelhead: What do we really know?, p. 337-353. In H.L., Jr. Schramm and R.G. Piper [ed.] Uses and Effects of Cultured Fishes in Aquatic Ecosystems. American Fisheries Society Symposium 15. American Fisheries Society, Bethesda, Maryland.
- Cederholm, C.J. et al. 1999. Pacific salmon carcasses: Essential contributions of nutrients and energy for aquatic and terrestrial ecosystems. Fisheries 24 (10): 6-15.
- Chapman, D., and K. Witty. 1993. Habitat of weak salmon stocks in the Snake River basin and feasible recovery measures. Report to the Bonneville Power Administration, DOE/BP-99654-1, Portland, Oregon.
- Congleton, J.L., and 10 coauthors. 1995. Evaluation procedures for collection, bypass, and downstream passage of outmigrating salmonids. Draft annual report for 1995, MPE-96-10.
- Dawley, E.M., R.D. Ledgerwood, T.H. Blahm, C.W. Sims, J.T. Durkin, R.A. Kirn, A.E. Rankis, G.E. Monan, and F.J. Ossiander. 1986. Migrational characteristics, biological observations, and relative survival of juvenile salmonids entering the Columbia River estuary, 1966-1983. 1985 Final Report. Bonneville Power Administration and National Marine Fisheries Service, Portland, Oregon.

DRAFT

- Elliott, D., and R. Pascho. 1994. Juvenile fish transportation: impact of bacterial kidney disease on survival of spring/summer chinook salmon stocks. 1992. Annual report of the National Biological Survey to the U.S. Army Corps of Engineers, Walla Walla, Washington.
- Elliot, D.G., R.J. Pascho, L.M. Jackson, G.M. Mathews, and J.R. Harmon. 1997. *Renibacterium salmoninarum* in spring-summer chinook salmon smolts at dams on the Columbia and Snake Rivers. *J. Aquat. Animal Health* 9: 114-126.
- Emlen, J.M., R.R. Reisenbichler, A.M. McGie, and T.E. Nickelson. 1990. Density-dependence at sea for coho salmon (*Oncorhynchus kisutch*). *Can. J. Fish. Aquat. Sci.* 47: 1765-1772.
- Enhancement Planning Team. 1986. Salmon and steelhead enhancement plan for the Washington and Columbia River conservation area. Vol. 1. Preliminary review draft.
- Goede, R. 1986. Management considerations in stocking of diseased or carrier fish. Pages 349-356 in R.H. Stroud, editor. *Fish Culture in fisheries management*. American Fisheries Society, Bethesda, Maryland.
- Groot, C. and L. Margolis. 1991. *Pacific salmon life histories*. UBC Press, University of British Columbia, Vancouver B.C.
- IHOT (Integrated Hatchery Operations Team). 1995. Policy and procedures for Columbia Basin anadromous salmonid hatcheries. Annual report 1994 to the Bonneville Power Administration, Portland Oregon. Project # 92-043.
- IHOT (Integrated Hatchery Operations Team). 1996. Operation plans for anadromous fish production facilities in the Columbia River Basin, Volume III-Washington. Annual report 1995 to the Bonneville Power Administration, Portland, Oregon. Project 92-043.
- Isaak, D.J., and T.C. Bjornn. 1996. Movement of northern squawfish in the tailrace of a lower Snake River dam relative to the migration of juvenile anadromous salmonids. *Transactions of the American Fisheries Society* 125: 780-793.
- Mantua, N.J., S.R. Hare, Y. Zhang, J.M. Wallace, and R.C. Francis. 1997. A Pacific interdecadal climate oscillation with impacts on salmon production. *Bull. Am. Meteorol. Soc.* 78: 1069-1079.
- McNeil, W.J. and D.C. Himsworth. 1980. *Salmonid ecosystems of the North Pacific*. Oregon State University Press and Oregon State University Sea Grant College Program, Corvallis, Oregon.

DRAFT

- Montgomery Watson. 1997a. Hatchery evaluation report. Little White Salmon NFH - coho. Report to the Bonneville Power Administration, Portland Oregon. Project # 95-2. Montgomery Watson, Bellevue, Washington.
- Montgomery Watson. 1997b. Hatchery evaluation report. Willard NFH - coho. Report to the Bonneville Power Administration, Portland Oregon. Project # 95-2. Montgomery Watson, Bellevue, Washington.
- Muir, W.D., and R.L. Emmett. 1988. Food habits of migrating salmonid smolts passing Bonneville Dam in the Columbia River, 1984. *Regulated River* 2: 1-10.
- Muir, W.D., A.E. Giorgi, and T.C. Coley. 1994. Behavioral and Physiological Changes in Yearling Chinook Salmon During Hatchery Residence and Downstream Migration. *Aquaculture* 127: 69-82.
- NMFS (National Marine Fisheries Service). 1997. Investigation of Scientific Information on the Impacts of California Sea Lions and Pacific Harbor Seals on Salmonids and on the Coastal Ecosystems of Washington, Oregon, and California. U.S. Dep. Commer., NOAA Tech. Memo. NMFS-NWFSC-28, 172 p.
- NMFS (National Marine Fisheries Service). 1999a. Biological Opinion on Harvest in the Columbia River Basin, Endangered Species Act - Section 7 Consultation.
- NMFS (National Marine Fisheries Service). 1999b. Biological Assessment for Mitchell Act Hatchery Operations. Hatcheries and Inland Fisheries Branch, Portland, Oregon.
- NMFS (National Marine Fisheries Service). 1999c. Biological Opinion on Artificial Propagation in the Columbia River Basin, Endangered Species Act - Section 7 Consultation.
- Nelson, W.R. and J. Bodle. 1990. Ninety years of salmon culture at Little White Salmon National Fish Hatchery. U.S. Fish & Wildlife Service, Washington, D.C.
- Olla, B.L., M.W. Davis and C.H. Ryer. 1993. Behavioral deficits of hatchery-reared Pacific salmon: potential effects on survival following release, p. 19. *In* D.S. Danielssen and E. Moksness [ed.] *Proc. Int. Symp. on Sea Ranching of Cod and Other Marine Animals*, held in Bergen, Norway, June 15-18, 1993. Institute of Marine Research, Bergen, Norway.
- Park, D.L. 1993. Effects of marine mammals on Columbia River salmon listed under the Endangered Species Act, DOE/BP-99654-3. Bonneville Power Administration. Portland, Oregon.
- Pastor, S.M. 1999. Annual coded wire program. Annual report 1998 to the Bonneville Power Administration, project 89-065, U.S. Fish and Wildlife Service, Vancouver, Washington.

DRAFT

- Piggins, D.J., and C.P.R. Mills. 1985. Comparative aspects of the biology of naturally-produced and hatchery-reared Atlantic salmon smolts *Salmo salar*. *Aquaculture* 45: 321-334.
- Roby, D. et al. 1997. Avian predation on juvenile salmonids in the lower Columbia River. 1997 Annual Report submitted to Bonneville Power Administration and U.S. Army Corps of Engineers.
- Ruggerone, G.T. 1986. Consumption of migrating juvenile salmonids by gull foraging below a Columbia River dam. *Trans. Am. Fish. Soc.* 115: 736-742.
- Sager, P.M., and G.J. Glova. 1988. Diet feeding periodictiy, daily ration and prey selection of a riverine population of juvenile chinook salmon, *Oncorhynchus tshawytscha*. *J. Fish Biol.* 33: 643-653.
- Salonius, K., and G.K. Iwama. 1993. Effects of Early Rearing Environment on Stress Response, Immune Function, and Disease Resistance in Juvenile Coho (*Oncorhynchus kisutch*) and Chinook Salmon (*O. tshawytscha*). *Canadian Journal of Fisheries and Aquatic Sciences* 50: 759-766.
- Schreck, C.B., A.G. Maule and S.L. Kaattari. 1993. Stress and disease resistance, p. 170-175. *In* J.F. Muir and R.J. Roberts [ed.] *Recent Advances in Aquaculture*, Vol. 4. Blackwell Scientific Publications, Oxford, UK.
- Simenstad, C., K. Fresh, and E. Salo. 1982. The role of Puget Sound and Washington coastal estuaries in the life history of Pacific Salmon: an unappreciated function. Pages 343-364 *in* V. Kennedy, editor. *Estuarine comparisons*. Academic Press, New York.
- Steward, R., and T. Bjornn. 1990. Supplementation of salmon and steelhead stocks with hatchery fish: a synthesis of published literature. Tech. Report 90-1. Part 2 *in* W.H. Miller, editor. *Analysis of Salmon and Steelhead Supplementation*. Bonneville Power Administration, Portland, Oregon. U.S. Fish and Wildlife, Dworshak Fisheries Assistance Office, Idaho.
- STT (Salmon Technical Team). 1999. Preseason report III. Analysis of council adopted management measures for 1999 ocean salmon fisheries. Pacific Fishery Management Council, Portland, Oregon.
- SWIG (Species Interaction Work Group). 1984. Evaluation of potential interaction effects in the planning and selection of salmonid enhancement projects. J. Rensel chairman and K. Fresh editor. Report prepared for the Enhancement Planning Team for implementation of the Salmon and Steelhead Conservation and Enhancement Act of 1980. Washington Department of Fish and Wildlife Olympia, Washington. 80pp.

DRAFT

Trotter, P.C. 1997. Sea-run cutthroat trout: life history profile. Pages 7-15 in J.D. Hall, P.A. Bisson, and R.E. Gresswell, editors. Sea-run cutthroat trout: biology, management, and future conservation. Oregon Chapter, American Fisheries Society, Corvallis, Oregon.

USFWS (U.S. Fish and Wildlife Service). 1994. Biological Assessments for operation of U.S. Fish and Wildlife Service operated or funded hatcheries in the Columbia River Basin in 1995-1998. Submitted to National Marine Fisheries Service under cover letter, dated August 2, 1994, from Bill Shake Acting USFWS Regional Director to Brian Brown, NMFS.

USFWS (U.S. Fish & Wildlife Service). 1987. Station development plan: Little White Salmon/Willard National Fish Hatchery Complex Little White Salmon/Willard NFH Complex. Division of Engineering, Portland, OR.

WDFW (Washington Department of Fish and Wildlife) and Confederated Tribes and Bands of the Yakima Indian Nation. 1990. Little White Salmon subbasin salmon and steelhead production plan. Columbia Basin System Planning. Report for the Northwest Power Planning Council, Portland , Oregon.

WDFW (Washington Department of Fish and Wildlife). 1997. Washington State salmonid stock inventory: appendix bull trout and dolly varden. Washington Department of Fish and Wildlife Olympia, Washington.